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Antibiotics as Major Disruptors of Gut Microbiota

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OPEN ACCESS

Edited by:
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Benjamin Mullan,
Imperial College London,
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Specialty section:

This article was submitted to
Microbiome in Health and Disease,
a section of the journal
*Frontiers in Cellular
and Infection Microbiology*

Received: 17 June 2020

Accepted: 29 October 2020

Published: 24 November 2020

Citation:
Ramirez J, Guarner F,
Bustos Fernandez L, Maruy A,
Sdepanian VL and Cohen H
(2020) Antibiotics as Major
Disruptors of Gut Microbiota.
Front. Cell. Infect. Microbiol. 10:572912.
doi: 10.3389/fcimb.2020.572912

INTRODUCTION

Scientific advances made in recent decades have led to an increased recognition of the role of the human gut microbiota in health and disease (Ananthakrishnan et al., 2019). Prior to that, little research had been conducted into the non-pathogenic microorganisms that inhabit the gastrointestinal tract (Guarner, 2012). Because most of these microorganisms cannot be cultured, they remained largely unexplored before the advent of molecular techniques. Ample experimental and clinical evidence now shows that gut microorganisms are required for the optimal functioning of the human body (Guarner, 2015).

In 1885, Louis Pasteur hypothesized that animals raised in sterile conditions would not be able to survive (Pasteur, 1885). Bernard S. Westmann and his team proved Pasteur's hypothesis wrong when they developed methods for breeding animals in germ-free conditions (Westmann, 1981).

However, they discovered that germ-free animals required large quantities of nutrient-rich food, yet



Microbiology Guide to Interpreting Minimum Inhibitory Concentration (MIC)

Historically, most *in vitro* susceptibility testing was performed by disk diffusion (Kirby-Bauer) method. The size of the growth-free zone determined whether the bacterium was considered to be susceptible, resistant or intermediate to a particular antibiotic.

While used as a guide to select an effective antibiotic, Kirby-Bauer testing could not tell the clinician the exact concentration of antibiotic needed to achieve a therapeutic result. Now, by a quantitative method of susceptibility testing known as the minimum inhibitory concentration (MIC), the precise concentration of antibiotic required to inhibit growth of a pathogen can be determined.

Your iDEXX microbiology results will show the identity of the organism and the appropriate antibiotic sensitivity pattern against each organism. Most antibiograms will include MICs in order to determine the most effective antibiotic that will result in effective treatment.

This guide provides a detailed explanation of the following concepts important in implementing the MIC:

- The MIC number is the lowest concentration ($\mu\text{g}/\text{mL}$) of an antibiotic that inhibits the growth of a given strain of bacteria. (See the "What Is an MIC?" section.)
- An MIC number for one antibiotic CANNOT be compared to the MIC number for another antibiotic. (See the "How Are MICs Used?" section.)
- The choice of antibiotic should be based on the MIC number, the site of infection and an antibiotic's breakpoint. Consider safety, ease of use and cost when determining the optimum antibiotic.

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BACTERIAL EMPIRE
2019, VOL. 2, NO. 4, 99-102

SCICELL

REGULAR ARTICLE

ANTIBIOTICS PRODUCING BACTERIA ISOLATED FROM FARMLANDS

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ABSTRACT

The need for new antibiotics has been highlighted recently with the increasing pace of emergence of drug resistance pathogens. Emerging strains of bacteria resistant to most advanced antibiotics have become issues of very important public health concern. Modification of existing antibiotics with the addition of side chains or other chemical group and protein-based drug targeting have been the preferred method of drug development at the corporate level in recent years. In this regard, soil samples were collected from farmland located in Oyo state, South Western part of Nigeria. Soil samples were collected from 16 selected farmland soils were characterized and assayed for antibiotic production and activity against a wide range of bacteria including *Klebsiella pneumoniae*, *Serratia marcescens*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Staphylococcus aureus*, *Bacillus megaterium* and *Enterococcus faecalis*. The extracts of the isolate obtained from *Streptomyces albulans*, *Streptomyces albus*, *Bacillus megaterium* and *Bacillus subtilis* showed activities against minimum of 3 and maximum of 4 of the 7 tested bacteria. Inhibition zones were found to range between 2.0 – 25.0 mm diameter at a concentration of 1ml. The minimum inhibitory concentration (MIC) of the crude extracts against the tested organisms ranged from 50% and above.

Bacillus megaterium and *Streptomyces albulans* were able to inhibit all the pathogenic bacteria, while *X. campestris* was unable to inhibit *Pseudomonas vulgaris* and *Staphylococcus aureus*, and *B. subtilis* was unable to inhibit *Enterococcus faecalis*.

Keywords: Antibiotics, *Bacillus*, Bacteria, Soil, Streptomyces

INTRODUCTION

The term soil refers to the outer loose material of the earth crust (Certiñí and Ugolini, 2013). Living portion of the soil body includes small animals and microorganisms such as fungi and bacteria, but it is generally considered that it's microorganisms that play the important role in soil (Arfuan et al., 2010).

Antimicrobials bacteria isolated from farmland have their ability to produce antibiotics and the *Streptomyces* is the dominant group (Pérez et al., 2014). The genus *Streptomyces* is responsible for the formation of more than 60% of known antibiotics (Gad et al., 2015; Rashad et al., 2015). Another important species of bacteria known to produce antibiotics is the *Bacillus* spp. Antibiotics are low molecular weight organic compounds produced by microorganisms as secondary metabolites in nature or microorganisms that live in soil (Jain et al., 2007).

Resistance of pathogenic bacteria has become a major health concern, many Gram positive bacteria and Gram negative opportunistic pathogens were becoming resistant to virtually every clinically available drugs (Rodríguez et al., 2014). The use of antimicrobial drugs for prophylactic or therapeutic purposes in humans and animals has increased over time due to the favorable environmental pressure favouring the survival and spread of resistant organisms. Selective pressure favouring the survival and spread of vancomycin-resistant enterococci (VRE) was the consequence of the use of antibiotics in food and agricultural practices. Vancomycin-resistance is often associated with multiple-drug resistance (Chang et al., 2015).

Another cause of concern is the Gram-negative antibiotic-resistant opportunistic pathogens. These bacteria, like *Pseudomonas aeruginosa*, are common environmental organisms, which act as opportunistic pathogens in clinical cases where the defense system for patient is compromised (Samjar et al., 2016). For instance, over 80 % of cystic fibrosis (CF) patients become chronically infected with *P. aeruginosa*. In addition, other opportunistic antibiotic resistant bacteria such as *Acinetobacter*, *Candida* and *Stenotrophomonas maltophilia* are emerging as opportunistic pathogens (Cheng et al., 2013). The appearance of multi-resistant pathogenic strains have caused a therapeutic problem of enormous proportions (Barbry et al., 2001). For instance, they cause substantial morbidity and mortality especially among the elderly and immunocompetent individuals. The incidence of antibiotic resistance problems of antibiotic resistance are developing at an alarming rate. Thus, new therapeutic drugs and approaches are needed to improve the management of these diseases and overcome these problems (Taylor et al., 2002).

Hence, intensive search for new antibiotics has become imperative worldwide especially from natural sources such as soil which is known as the greatest source of antibiotics. New microbial metabolites are permanently needed due not only to the increase in resistant pathogens, but also to the evolution of novel diseases and toxicity of currently used compounds (Paragone et al., 2008).

This study aims at exploring the antibiotics production potentials of some indigenous soil bacteria isolated from farmlands in some selected locations in Oyo and Osun state, South Western part of Nigeria. The objectives include: (i) to isolate antibiotic producing bacteria from farmland (ii) to characterize to species level the isolated bacteria (iii) to determine and estimate the antimicrobial efficiency of each of the antibiotics produced by the bacteria.

MATERIALS AND METHOD

Study area

The soil samples were collected from farmlands located in Ibadan, Ago - Ife, and Ilorin. Seven samples were collected from individual farmlands in Ibadan which is the capital of Oyo state, while five samples were collected from government farmland located in Ago - Ife, and four other samples were collected from IITA research station located in Ilorin, both located in Osun state.

Media used

The media used were nutrient agar, potato starch casein agar, starch casein agar, and Mac Conkey agar. They were prepared according to the manufacturer's instruction. The media were left to cool to about 45°C after sterilization before being poured aseptically into the Petri dish.

Serial dilution of soil samples: The soil samples were labeled according to the area of collection. Then 1g was weighed from each soil samples for serial dilution where the 10^0 to 10^7 dilutions were plated.

Plating of the samples

The diluted soil samples were inoculated onto the appropriate agar medium using the streaking method, and they were labeled accordingly. Nutrient agar was used for the isolation of *Bacillus* spp. and *Streptomyces* spp. and Mac Conkey agar for *Escherichia coli* and *Klebsiella pneumoniae*. After inoculation, the plates were incubated at 37°C over night for bacteria, and while in the case of *Escherichia coli* incubation lasted for 4 days at a temperature of 30°C before morphological changes can be seen.

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Antibiotics Susceptibility Test

- Two tests can be used: dilution series test and agar diffusion test
- Dilution series test → antibiotic agents are prepared in the nutrient medium, inoculated with organism, incubated, then the lowest growth-inhibiting concentration level are determined

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1. Abraham, E. P., and E. Chain. 1940. An enzyme from bacteria able to destroy penicillin. *Rev. Infect. Dis.* 10:677-678. [PubMed] [Google Scholar]2. Alekshun, M. N., and S. B. Levy. 2007. Molecular mechanisms of antibacterial multidrug resistance. *Cell* 128:1037-1050. [PubMed] [Google Scholar]3. Allen, H. K., J. Donato, H. H. Wang, K. A. Cloud-Hansen, E. Davies, and J. Handelsman. 2007. The antibiotic resistance genes in natural environments. *Nat. Rev. Microbiol.* 5:243-251. [PubMed] [Google Scholar]4. Allen, H. K., L. A. Moe, J. Rodbumr, A. Gaarder, and J. Handelsman. 2009. Functional metagenomics reveals diverse beta-lactamases in a remote Alaskan soil. *ISME J.* 3:243-251. [PubMed] [Google Scholar]5. Allou, N., E. Cambau, L. Massias, F. Chau, and B. Fantic. 2009. Impact of low-level resistance to fluoroquinolones due to *gyrA* mutation on ciprofloxacin bactericidal activity in a murine model of *Escherichia coli* urinary tract infection. *Antimicrob. Agents Chemother.* 53:4292-4297. [PMC free article] [PubMed] [Google Scholar]6. American Academy of Microbiology. 2009. Antibiotic resistance: an ecological perspective on an old problem. Based on a colloquium held in the Fondation Mérieux Conference Center in Annecy, France, 12 to 14 October 2008. ASM Press, Washington, DC. 7. American Academy of Microbiology. 2005. Vaccine development: current status and future needs. Based on a colloquium held in Washington, DC, 4 to 6 March 2005. ASM Press, Washington, DC. 8. Aminov, R. I. 2009. The role of antibiotics and antibiotic resistance in nature. *Environ. Microbiol.* 11:2970-2988. [PubMed] [Google Scholar]9. Aminov, R. I., and R. I. Mackie. 2007. Evolution and ecology of antibiotic resistance genes. *FEMS Microbiol. Lett.* 271:147-161. [PubMed] [Google Scholar]10. Andersson, D. I. 2006. The biological cost of mutational antibiotic resistance: some practical conclusions? *Curr. Opin. Microbiol.* 9:461-465. [PubMed] [Google Scholar]11. Balaban, N., T. Goldkorn, R. N. Hanan, L. B. Dang, S. Scott, R. M. Ridgley, A. Rasouly, S. C. Wright, J. W. Lerrick, R. Rasouly, and J. P. Carlson. 2009. A paradigm of virulence as a target for vaccine and therapy against *Staphylococcus aureus*. *Science* 280:438. [PubMed] [Google Scholar]12. Baltz, R. H. 2006. Marcel Fabre Roundtable: is our starvation, scott or r. m. ridgley, a paradigm of virulence as a target for vaccine and therapy against *Staphylococcus aureus*. *Science* 280:438. [PubMed] [Google Scholar]13. Barbe, F., J. L. Martínez, and R. Cantón. 2008. Antibiotics and antibiotic resistance in water environments. *Curr. Opin. Biotechnol.* 19:260-265. [PubMed] [Google Scholar]14. Barbe, F., D. Vallenet, N. Pöhlkecheen, A. Kreimeyer, S. Ozras, L. Labeyrie, S. Crivellari, C. Robert, S. Duprat, P. Wincker, L. N. Ormiston, J. Weissenbach, P. Marliere, G. N. Cohen, and M. Médigue. 2004. Unique features revealed by the genome sequence of *Acinetobacter* sp. ADP1, a versatile and naturally transforming competent bacterium. *Nucleic Acids Res.* 32:766-779. [PMC free article] [PubMed] [Google Scholar]15. Barlow, M., and J. G. Hall. 2002. Phylogenetic analysis shows that the OXV beta-lactamase genes have been transferred millions of years ago. *Mol. Evol.* 19:311-321. [PubMed] [Google Scholar]16. Bartolucci, H., R. Rodriguez, C. Fernandez, A. Mantilla, and J. Bartolucci. 2009. Antibiotic resistance in *Enterococcus faecalis* from Argentina: diversity of resistance genes and distribution of plasmid-mediated resistance. *Natl. Acad. Sci. U. S. A.* 106:4896-4901. [PMC free article] [PubMed] [Google Scholar]17. Benveniste, R., and J. Davies. 1973. Aminoglycoside antibiotic-inactivating enzymes: actinomycetes similar to those present in clinical isolates of antibiotic-resistant bacteria. *Proc. Natl. Acad. Sci. U. S. A.* 70:2276-2280. [PMC free article] [PubMed] [Google Scholar]18. Benveniste, R., and J. Davies. 1973. Aminoglycoside antibiotic-inactivating enzymes: actinomycetes similar to those present in clinical isolates of antibiotic-resistant bacteria. *Proc. Natl. Acad. Sci. U. S. A.* 70:2276-2280. [PMC free article] [PubMed] [Google Scholar]19. Benveniste, R., and J. Davies. 1973. Aminoglycoside antibiotic-inactivating enzymes: actinomycetes similar to those present in clinical isolates of antibiotic-resistant bacteria. *Proc. Natl. Acad. Sci. U. S. A.* 70:2276-2280. [PMC free article] [PubMed] [Google Scholar]20. Boucher, H. W., G. H. Talbot, J. S. Bradley, J. E. Edwards, D. Gilbert, L. B. Rice, M. Scheckl, B. Spellberg, and J. Bartlett. 2009. Bad bugs, no drugs: an update from the Infectious Diseases Society of America. *Clin. Infect. Dis.* 48:1-12. [PubMed] [Google Scholar]21. Boucher, Y., M. Labbate, J. E. Koening, and H. W. Stokes. 2007. Integrating mobileable platforms that promote genetic diversity in bacteria. *Trends Microbiol.* 15:301-309. [PubMed] [Google Scholar]22. Bouvier, M., M. Duco-Galand, C. Loot, B. Bikard, and D. Mazel. 2009. Structural features of single-stranded integron cassette attC sites and their role in selection. *PLoS Genet.* 5:e1000032. [PubMed] [Google Scholar]23. Brüssow, H., and R. E. Hancock. 2008. Using microarray gene signatures to elucidate mechanisms of antibiotic action and resistance. *Drug Discov. Today* 10:1245-1252. [PubMed] [Google Scholar]24. Brüssow, H., M. Arthur, and P. Courvalin. 1988. Evidence for natural gene transfer from gram-positive cocci to *Escherichia coli*. *J. Bacteriol.* 170:1739-1745. [PMC free article] [PubMed] [Google Scholar]25. Brochet, M., E. Couvè, M. Zouine, C. Poyart, and P. Glaser. 2008. A naturally occurring gene amplification leading to sulfonamide and trimethoprim resistance in *Streptococcus agalactiae*. *J. Bacteriol.* 190:672-680. [PMC free article] [PubMed] [Google Scholar]26. Brötz-Oesterhert, H., and N. A. Brunner. 2008. How many modes of action should an antibiotic have? *Curr. Opin. Pharmacol.* 8:564-573. [PubMed] [Google Scholar]27. Bryskier, A. (ed.). 2005. Antimicrobial agents: antibiotics and antifungals. ASM Press, Washington, DC. 28. Bush, K., and G. A. Jacoby. 2010. Updated functional classification of beta-lactamases. *Antimicrob. Agents Chemother.* 54:969-976. [PMC free article] [PubMed] [Google Scholar]29. Bushman, F. 2002. Lateral DNA transfer. *Cold Spring Harbor* Press, Cold Spring Harbor, NY. 30. Canton, R. 2009. Antibiotic resistance genes from the environment: a perspective through newly identified antibiotic resistance mechanisms in the clinical setting. *Clin. Microbiol. Infect.* 15(Suppl. 1):S1-15. [PubMed] [Google Scholar]31. Carisson, G., S. Orn, and D. G. J. Larsson. 2009. Effluent from bulk drug production is toxic to aquatic vertebrates. *Environ. Res.* 109:105-118. [PubMed] [Google Scholar]32. Chater, K. F., and C. Bruton. 1985. Resistance, regulatory and production genes for the antibiotic methylenomycin are clustered. *EMBO J.* 4:229-241. [PMC free article] [PubMed] [Google Scholar]33. Chee-Sanford, J. C., R. I. Mackie, S. Kolke, J. G. Krackap, Y.-F. Lin, A. C. Vannarelli, S. Maxwell, and R. I. Aminov. 2009. Fate and transport of antibiotic residues and antibiotic resistance genes following land application of manure waste. *J. Environ. Qual.* 38:1086-1106. [PubMed] [Google Scholar]34. Couce, A., and J. Blazquez. 2009. Side effects of antibiotics on genetic variability. *FEMS Microbiol. Rev.* 33:531-538. [PubMed] [Google Scholar]35. Dantas, G., M. O. A. Sommer, R. D. Olujohungbe, and G. M. Church. 2008. Bacteria subsisting on antibiotics. *Science* 320:100-103. [PubMed] [Google Scholar]36. Da Re, S., F. Garnier, E. Guerin, S. Campoy, F. Deniz, and M. C. Ploy. 2009. The SOS response promotes qnrB quinolone-resistance determinant expression. *EMBO Rep.* 10:929-933. [PMC free article] [PubMed] [Google Scholar]37. Datta, N., and V. M. Hughes. 1983. Plasmids of the same Inc groups in enterobacteria before and after the use of antibiotics. *Science* 203:616-617. [PubMed] [Google Scholar]38. Davies, J. 1995. Vicious circles: looking back at resistance plasmids. *Genetics* 139:1465-1468. [PMC free article] [PubMed] [Google Scholar]39. Davies, J. 1990. What are antibiotics? Archaic functions for modern activities. *Mol. Microbiol.* 4:1227-1232. [PubMed] [Google Scholar]40. DeAngelis, D., and G. M. Church. 2008. Bacteria subsisting on antibiotics. *Science* 320:100-103. [PubMed] [Google Scholar]41. Davies, J., G. B. Spiegelman, and G. Yim. 2006. The world of subinhibitory antibiotic concentrations. *Curr. Opin. Microbiol.* 9:1-9. [PubMed] [Google Scholar]42. D'Costa, V. M., E. Griffiths, and G. D. Wright. 2007. Reemergence of antibiotic-resistant *Staphylococcus aureus* in the genetics era. *J. Clin. Invest.* 119:2464-2474. [PMC free article] [PubMed] [Google Scholar]43. D'Costa, V. M., K. M. McGrann, D. W. Hughes, and G. D. Wright. 2006. Sampling the antibiotic resistance genome. *Science* 311:374-377. [PubMed] [Google Scholar]44. Depardieu, F., R. F. de la Poerier, and G. F. Chambers. 2009. How many modes of action should an antibiotic have? *Curr. Opin. Microbiol.* 10:481-489. [PubMed] [Google Scholar]45. Desain, A. L., and S. Sanchez-Alberola. 2009. Microbial drug discovery: 80 years of progress. *J. Antibiot.* (Tokyo) 62:5-16. [PMC free article] [PubMed] [Google Scholar]46. Di Padova, F., I. Podglajen, R. Leclercq, E. Collatz, and P. Courvalin. 2007. Modes and modulations of antibiotic resistance gene expression. *Clin. Microbiol. Rev.* 20:79-114. [PMC free article] [PubMed] [Google Scholar]47. De Vincenti, S. J., and C. Viola. 2006. Delays of an advisory committee regarding priorities, sources, and methods for collecting animal antimicrobial use data in the United States. *Prev. Vet. Med.* 73:133-151. [PubMed] [Google Scholar]48. Dioniso, F., I. Matic, M. Radman, O. R. Roje, and F. Taddei. 2002. Plasmids spread very fast in heterogeneous bacterial communities. *Genetics* 162:1525. [PMC free article] [PubMed] [Google Scholar]49. Doyle, M. P. 2006. Antimicrobial resistance: implications for the food system. *Compr. Rev. Food Sci. Food Saf.* 5:71-137. [PubMed] [Google Scholar]50. Enright, M. C., D. A. Robinson, G. Rodriguez, E. J. Feil, H. Grundmann, and G. B. Spratt. 2002. The evolutionary history of methicillin-resistant *Staphylococcus aureus* (MRSA). *Proc. Natl. Acad. Sci. U. S. A.* 99:7687-7692. [PubMed] [Google Scholar]51. Fajardo, A., N. Martinez-Martin, M. Mercadillo, C. Galan, B. Chysells, S. Matthijs, P. Cornelis, L. Weihmann, B. Tummel, F. Baquero, and J. L. Martinez. 2008. The neglected intrinsic resistance of bacterial pathogens. *PLoS One* 1:1496-1502. [PubMed] [Google Scholar]52. Fajardo, A., N. Martinez-Martin, M. Mercadillo, C. Galan, B. Chysells, S. Matthijs, P. Cornelis, L. Weihmann, B. Tummel, F. Baquero, and J. L. Martinez. 2008. The production and role of antibiotics in soil. *J. Antibiot.* (Tokyo) 61:987-1000. [PubMed] [Google Scholar]53. Fenton, J. J., H. Harsch, and D. Klein. 1973. Production of volatile nitrogenous compounds from the degradation of streptomycin by *Pseudomonas maltophilia*. *J. Bacteriol.* 116:1267-1272. [PubMed] [Google Scholar]54. Fick, J., H. Soderstrom, R. H. Lindberg, C. Phan, M. Tykland, and G. J. Larsson. 2009. Contamination of surface, ground, and drinking water from pharmaceutical production. *Environ. Toxicol. Chem.* 28:2252-2257. [PubMed] [Google Scholar]55. Finland, J. 1979. Emergence of antibiotic resistance in hospitals, 1935-1975. *Rev. Infect. Dis.* 1:4-22. [PubMed] [Google Scholar]56. Finlay, B. B., and R. E. Hancock. 2004. Can innate immunity be enhanced to treat microbial infections? *Nat. Rev. Microbiol.* 2:497-504. [PubMed] [Google Scholar]57. Forsman, M., B. Häggström, L. Lindgren, and B. Jaurin. 1990. Molecular analysis of β -lactamases from four species of *Streptomyces*: comparison of amino acid sequences with those of other β -lactamases. *Microbiology* 136:589-598. [PubMed] [Google Scholar]58. Funnell, B. E., B. E. Phillips, and G. J. Phillips (ed.). 2004. *Plasmid biology*. ASM Press, Washington, DC. 59. Gale, E. F., E. Cundliffe, P. E. Reynolds, M. H. Richmond, and M. J. Waring (ed.). 1981. The molecular basis of antibiotic action, 2nd ed. John Wiley, Chichester, United Kingdom. 60. Gillings, M. Y., Boucher, M. Labbate, A. Holmes, S. K. Krishnan, M. Holley, and H. W. Stokes. 2008. The evolution of class I integrons and the rise of antibiotic resistance. *J. Bacteriol.* 190:5095-5100. [PMC free article] [PubMed] [Google Scholar]61. Gillings, M. R., M. P. Holley, and H. W. Stokes. 2009. Evidence for dynamic exchange of *qac* gene cassettes between class I integrons and other integrons in freshwater biofilms. *FEMS Microbiol. Lett.* 276:282-288. [PubMed] [Google Scholar]62. Gillings, M. R., S. Krishnan, P. J. Warden, and S. A. Hardwick. 2008. Recovery of diverse genes for class I integron-integrases from environmental DNA samples. *FEMS Microbiol. Lett.* 287:286-291. [PubMed] [Google Scholar]63. Ginidowski, M. 2008. Evolution of extended-spectrum beta-lactamases by mutation. *Clin. Microbiol. Infect.* 14(Suppl. 1):11-32. [PubMed] [Google Scholar]64. Gomez, M. J., and A. A. Neyfakh. 2006. Genes involved in intrinsic antibiotic resistance of *Acinetobacter baumannii*. *Antimicrob. Agents Chemother.* 50:3562-3567. [PMC free article] [PubMed] [Google Scholar]65. Gottlieb, D. 1976. The production and role of antibiotics in soil. *J. Antibiot.* (Tokyo) 29:987-1000. [PubMed] [Google Scholar]66. Gutierrez, E., G. Cambray, N. Sanchez-Alberola, S. Campoy, I. Erill, S. Da Re, B. Gonzalez-Zorn, J. Barbé, C. Ploy, and D. Mazel. 2009. The SOS response controls integron recombination. *Science* 324:1034-1037. [PubMed] [Google Scholar]67. Hackett, J., and J. B. Podglajen, R. Leclercq, E. Collatz, and P. Courvalin. 2007. Delays of an advisory committee regarding priorities, sources, and methods for collecting animal antimicrobial use data in the United States. *Prev. Vet. Med.* 73:133-151. [PubMed] [Google Scholar]68. Hall, G. M., and R. E. Hancock. 2004. Can innate immunity be enhanced to treat microbial infections? *Nat. Rev. Microbiol.* 2:497-504. [PubMed] [Google Scholar]69. Hacker, J., and J. B. Kaper. 2000. Pathogenicity islands and the evolution of microbes. *Annu. Rev. Microbiol.* 51:641-679. [PubMed] [Google Scholar]70. Hakenbeck, R. 1998. Mosaic genes and their role in penicillin-resistant *Streptococcus pneumoniae*. *Electrophoresis* 19:597-601. [PubMed] [Google Scholar]71. Hawkey, P. M., and A. M. Jones. 2009. The changing epidemiology of resistance. *J. Antimicrob. Chemotherapy* 64(Suppl. 1):S1-10. [PubMed] [Google Scholar]72. Heijnen, P. 2009. The X-ray analysis of the structure of penicillin. *Adv. Sci.* 6:85-89. [PubMed] [Google Scholar]73. Holmes, A. J., M. R. Gillings, B. S. Nield, B. C. Mabbatt, K. M. Nevalainen, and J. R. Winstone. 2008. The evolution of class I integrons and the rise of antibiotic resistance. *J. Bacteriol.* 190:5095-5100. [PMC free article] [PubMed] [Google Scholar]74. Horwitz, A. M., J. T. Puky, R. D. Ridder, G. Dzolje-Danilovic, J. Degener, K. F. Kerrebijn, and M. F. Meijer. 1999. Ecology of *Pseudomonas aeruginosa* patients with cystic fibrosis. *J. Med. Microbiol.* 31:1124. [PubMed] [Google Scholar]75. Jacobson, J. B., and J. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]76. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]77. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]78. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]79. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]80. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]81. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]82. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]83. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]84. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]85. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]86. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]87. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]88. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]89. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]90. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]91. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]92. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]93. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]94. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]95. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]96. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]97. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]98. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]99. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]100. Janczewski, T., and J. B. P. Kelly. 2009. Antimicrobial resistance: exploring environmental diversity and its consequences. *Environ. Microbiol.* 11:2272-2277. [PubMed] [Google Scholar]101. Janczewski,

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